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Harnessing the phytochemistry and pharmacological roles of Andrographis paniculata

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Abstract: Herbs and natural product have extremely high potential as therapeutic agent without causing serious side effects on human body. In this review article the medicinal uses of Andrographis peniculata an annual herb found in Asian continent and other parts of world is being discussed along with its traditional uses, phytochemistry and pharmacology.

Key words: Andrographis peniculata, Traditional medicinal uses, phytochemistry, pharmacology **Introduction:**

A. paniculata, commonly known as King of Bitters or kalmegh, is an annual, branched, erect herb. It is native to peninsular India and Srilanka and is also distributed in different regions of Southeast Asia, China, America, West Indies and Christmas Island. It is cultivated because of its well known medicinal value and it is widely distributed since it grows well in most soil types¹. The aerial parts and roots of the plant have been widely used as traditional medicine in China, India, Thailand and other Southeast Asian countries to treat many maladies. Its common names are King of Bitters (English), Mahatikta (Sanskrit), Kiryato (Gujarati), Mahatita (Hindi), Kalmegh (Bengali)². It is also known as 'Bhui Neem' as the plant although being very small has similarity in taste and appearance as Neem (Azadirachta indica). A wide array of studies has been conducted by researchers, especially in Asia, following reports about the medicinal properties possessed by this plant mostly according to traditional medical practitioners in avurvedic medical system. Phytochemical studies have revealed that A. paniculata contains diverse compounds including diterpenoid lactones, flavonoids and miscellaneous compounds. It has been shown to possess wide spectrum of pharmacological properties^{3,4}.



Figure 1: A. Paniculata leaves

Figure 2: A. Paniculata flower

Traditional Medicinal Uses

The aerial parts, roots and whole plant of A. paniculata have been used for centuries in Asia as traditional medicine for the treatment of various ailments. It has been used by traditional medical practitioners for stomachaches, inflammation, pyrexia, and intermittent fevers⁵⁻⁸. The whole plant has been used for several applications such as antidote for snake-bite and poisonous stings of some insects, and to treat dyspepsia, influenza, dysentery, malaria and respiratory infections^{5,6}. The leaf

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extract is a traditional remedy for the treatment of infectious disease, fever causing diseases, colic pain, loss of appetite, irregular stools and diarrhea⁹. In Malaysia, a decoction of the aerial parts is used to treat common cold, hypertension, diabetes, cancer, malaria and snake bite¹⁰. It is an important constituent of at least 26 Ayurvedic formulas in Indian pharmacopoeia. In traditional Chinese medicine, it is seen as the cold-property herb used to rid the body of heat and fever and to dispel toxins from the body¹¹. In Ayurvedic medicinal system, tribals of Tamilnadu (India use this herb for a variety of ailments like dysmenorrhoea, leucorrhoea, pre-natal and post-natal care, complicated diseases such malaria, jaundice, gonorrhea and general ailments like wounds, cuts, boils and skin diseases¹²⁻¹⁴.

A. paniculata is being widely used by the Local Traditional Healers of Chhattisgarh also mainly in the treatment of Cough & Cold, Fever, Jaundice and Diabetes apart from other ailment treatments. The plant is generally used in combination with some other medicinal plants or their parts as a compound drug¹⁵.

Phytochemistry

A. paniculata has various compounds in its aerial parts and roots and these are often used in extracting its active principles. Diverse factors such as geographical region, harvest time and processing method account for the variability in its chemical content. Phytochemical tudies of *A. paniculata* has led to the isolation of various plant metabolites. Notable among these metabolites are the terpenoids (entalabdane diterpene lactones) which account for a large proportion of its components and therapeutic activities¹⁶⁻¹⁸. Other categories of compounds that have also been isolated include flavonoids (flavones), noriridoides, xanthones, polyphenols and trace and macro elements¹⁹.

Terpenoids

Diterpenoid lactones are the commonest terpenoid compounds isolated from *A. paniculata* (Table 1). Diterpenoids are distributed in and have been isolated from the aerial parts and roots of this plant. of the diterpenoids that have been identified and isolated from A. paniculata, andrographolide is the most prominent in occurrence and quantity. Andrographolide has a very bitter taste, and it is colourless and crystalline in appearance¹⁶ and was first isolated in pure form by Gorter in 1911. Dominant diterpenoids other than andrographolide which have been isolated mostly from the aerial parts of A. paniculata include deoxyandrographolide and neoandrographolide. These diterpenoids (Table 1) have been isolated by several workers¹⁷⁻¹⁸.

Flavonoids

Flavones are the major flavonoids that have been isolated from the aerial parts, roots and whole plant of *A. paniculata* (Table 1)^{19,20}.

Other compounds

Several other compounds (Table 1) have also been isolated, especially, from the roots of *A*. *paniculata*. Four xanthones were isolated from the roots identified as 1, 8 -dihydroxy- 3, 7 - dimethoxy-xanthone, 4, 8 -dihydroxy- 2, 7 -dimethoxy-xanthone, 1, 2 -dihydroxy- 6, 8 - dimethoxy- xanthone and 3, 7, 8 -trimethoxy- 1 -hydroxy-xanthone 21 . F ive rare noriridoids designated as andrographolide A - E, along with curvifloruside were isolated from the roots of A. paniculata²².

SN.	Compound	Plant part from where isolated	Compound type
1.	Andrographolide	Leaves/aerial	Diterpenoid
	r marographonae		lactone
2.	Neoandrographolide	Leaves/aerial	Diterpenoid
2.	reoundrographonide		lactone
3.	14-deoxyandrographolide	Aerial parts	Diterpenoid
5.	14 deoxyandrographonde		lactone
4.	Andrographoside	Leaves/aerial parts	Diterpene
ч.	Andrographoside	Leaves/ aerial parts	Diterpene
5.	14-deoxy-11, 12-	Aerial parts	Diterpenoid
	didehydroandrographolide		lactone
6.	5, 7, 2', 3'-	Whole plant	Flavonone
	tetramethoxyflavone		
7.	5-hydroxy-7, 2', 3'-	Whole plant	Flavone
	trimethoxy flavones		
8.	6'trimethoxyflavone	Root	Flavone
9.	5-hydroxy-7, 2', 7-O-	Root/aerial part	Flavone
	methyldihydrowogonin		
10.	7-O-methylwogonin	Root/aerial part/whole plant	Flavone
11.	Flavone-1, 2'methylether	Root/aerial part/whole plant	Flavone
12.	7-O-methylwogonin-5-	Root/aerial parts	Flavones
	glucoside		
13.	Flavone-1, 2'-O-glucoside	Root /aerial part/whole plant	Flavonoids
14.	5-hydroxy-7, 8, 2', 5'-	Whole plant	Flavonoids
	tetramethoxyflavone		

Pharmacology

The use of the different parts of *A. paniculata* plant in folk medicine, especially, in Asia led scientists to study its pharmacological properties to explore the use of this plant as a therapeutic agent in the remedy of various ailments. Several studies showed that this plant exhibited various biological activities such as anti-microbial, cytotoxicity, anti-protozoan, anti-inflammatory, anti-oxidant, immunostimulant, anti-diabetic, anti-infective, anti-angiogenic, hepato-protective, sex hormone modulatory, liver enzymes modulatory and insecticidal and toxicity activities and cardiovascular therapy.

Anti-microbial activity

Aqueous extract, andrographolides and arabinogalactan proteins isolated from the dried herb of *A*. *paniculata* were screened for anti-microbial activity. The result showed that the aqueous extract and arabinogalactan proteins have antibacterial activity against Bacillus subtilis (*B. subtilis*), *Escherichia coli* (*E. coli*), *Pseudomonas aeruginosa* while andrographolide was only active against *B. subtilis*. All three were also reported to possess anti-fungal activity against *Candida albicans*²³. Five rare noriridoides, andrographidoides A - E were screened for anti-bacterial activity against E. coli,

Staphylococcus aureus, Staphylococcus epidermidis, Pseudomonas aeruginosa and *B. subtilis*. None of the compounds showed any inhibitory activity (MIC>100 μ g/mL). Gentamycin, chloramphenicol and Ciprofloxacin were used as positive controls ²².

Anti-inflammatory/anti-allergic activity

The A. paniculata leaves combined aqueous and methanol extract showed significant improvement of lipopolysacharide inducer release of pro-inflamatory mediators (NO, IL-1ß and IL-6), inflammatory mediators (PGE 2 and TXB2) along with allergic mediators (LTB 4)²⁴. Different photochemicals: andrographolide, neoandrographolide, isoandrographolide, andrograpanin, 7-Omethylwogonin, skullcapflavone and 14 -deoxy- 11, 12 - didehydroandrographolide isolated from A. paniculata leaves were screened for in vitro anti-inflammatory and anti-allergic potential. It was observed that andrographolide, isoandrographolide, 7-O-methylwogonin and skullcapflavone-1 significantly inhibited inflammatory mediators NO and PGE2 release from lipopolysacharide (LPS) stimulated cultured macrophages. Andrographolide, isoandrographolide and 7- O-methylwogonin inhibited the production of IL-1 β in LPS stimulated macrophages. In addition IL - 6 production from LPS induced macrophages was significantly (P<0.01) inhibited by andrographolide, isoandrographolide and skullcapflavone-1 in a concentration dependent manner. The results also showed that andrographolide, isoandrographolide and skullcapflavone-1 significantly suppressed TXB 4 released in A 23187 activated HL - 60 promyelocytic leukemia cells. Andrographolide, dehydroandrographolide and neoandrographolide isolated from the aerial parts of A. paniculata exhibited anti-inflammatory effects by interfering with COX enzyme activity. Remarkable inhibition of COX-1 by andrographolide (30.1 µM) and dehydroandrographolide (28.5 µmol/ L) was found in ionophore A 23187-induced human platelets. Dehydroandrographolide (28.5 µmol/ L) and neoandrographolide (20.8 µmol/ L) strongly suppressed the LPS -stimulated COX - 2 activity in human blood and dehydroandrographolide modulated the level of LPS -induced TNF -α, IL - 6, IL -1β, and IL - 10 secretion in human blood in a concentration dependent manner. It indicates that dehydroandrographolide has the highest efficacy²⁵. Andrograpanin (15 - 90 μ mol/L) isolated from the ethanol extract of the leaves inhibited NO and pro-inflammatory cytokines in a dose dependent manner from lipopolysaccharide activated macrophages. Significant (p< 0.05) inhibition of NO was evident at a concentration of 30 µmol/L and at a concentration of 75 µmol/L. Andrograpanin almost completely inhibited NO production. Significant inhibition of proinflammatory cytokines was evident at a concentration of 1.5µmol/L and there was an almost complete inhibition at a concentration of 90 μ mol/ L²⁵.

In another study it was found that andrographolide has shown to have a strong anti-inflammatory activity. LPS-induced TNF- α and GM-CSF release from mouse peritoneal macrophages was inhibited by andrographolide in a concentration-dependent manner. The concentration of the drug producing 50% inhibition was 0.6 μ M for TNF- α and 3.3 μ M for GM-CSF. The maximal inhibition achieved (at 50 μ M) was 77% and 94%, respectively, for the two cytokines. In the *in vivo* study, intra-peritoneal treatment of ovalbumin-immunized and nasally-challenged mice with andrographolide significantly inhibited the elevation of bronchoalveolar fluid (BAF) levels of TNF- α and GM-CSF in a dose-dependent manner, with 30 mg/kg produced an inhibition of 92% and 65% of the cytokines and almost completely abolished the accumulation of lymphocytes and <u>eosinophils²⁶</u>.

The anti-allergic properties of the phytoconstituents was investigated on A23187 induced LTB4 production. The result showed 30.5% and 19.6% inhibition of LTB 4 production in A 23187 induced HL - 60 promyeolocytic leukemia cells at concentrations of 63 μ mol/L and 33.5 μ mol/L for

skullcapflavone and 7-O-methylwogonin respectively. The IC 50 value for the reference standard captopril was 48 μ mol/L. 7-O-methylwogonin was the only phytoconstituent that potently inhibited A23187 induced histamine release in RBL-2H3 rat basophil leukemic cells in a dose dependent manner ²⁷.

In another study diterpenes, andrographolide and neoandrographolide, isolated from the plant, were evaluated for antiallergic activity. These were tested for anti-PCA (Passive cutaneous anaphylaxis) and mast cell stabilizing activities in rats have shown significant anti-PCA activity²⁸.

Cytotoxicity

The methanol extract, petroleum ether, dichlomethane fraction and aqueous fraction of the methanol extract of A. peniculata were screened for anti-proliferation activity against HT-29 (colon cancer) cells. The methanol extract inhibited the proliferation of HT-29 cells by 50% at a concentration of 10 µg/mL. The petroleum ether and dichloromethane fractions inhibited proliferation of HT-29 cells with a GI50 value of 46 µg/mL and 10 µg/mL respectively. The aqueous extract did not inhibit the proliferation of HT-29 cells. The diterpenes adrographolide isolated from the dichlomethane fraction, inhibited the proliferation of all cancer cells screened. 14 -deoxyandrographolide showed moderate inhibition against the proliferation of two cancer cell out of the entire cell screened. 14-deoxy-11, 12-didehydroandrographolide did not inhibit the proliferation of any of the cancer cell line tested²⁷. These findings are in accordance with earlier reports, showing the cytotoxic activity of andrographolide against human epidermoid carcinoma and lymphocytic leukaemia cells¹⁸. The growth inhibitory activity on mouse myeloid leukemia cells by the methanol extract of the aerial parts of A. paniculata and some of the isolated compounds has also been reported²⁹. Andrographolide and its semisynthetic analogues namely 3. 19 isopropylideneandrographolide, 14-acetyl-3, -isopropylideneandrographolide 19 and 14 acetylandrographolide were screened in vitro for anti-tumor activity against MCF - 7 human breast cancer and HCT - 116 colon cancer cell lines. 19-isopropylideneandrographolide and 14 acetylandrographolide showed cytotoxic activity against the two cell lines tested and they were equally potent when compared to parent andrographolide. In a similar study at the national cancer institute in the USA, 19 -isopropylideneandrographolide and 14-acetylandrographolide were also screened and found to be cytotoxic against 60 human cancer cell lines³⁰. Xanthones isolated from the chloroform fraction of the roots were screened for cytotoxicity and the results showed that all the compounds have IC 50 values $>16 \mu g/m L$, which can be considered as non-cytotoxic as per WHO criteria²².

Antidiabetic effect:

A. paniculata has also been studied for its possible impact on the management of Diabetes Miletus in humans ${}^{31-34}$. Andrographolide is a potential bioactive phytochemical in *A. paniculata*, which possesses antidiabetic properties ${}^{31, 35-39}$. Oral andrographolide and *A. paniculata* lowered the blood glucose levels of streptozotocin-induced diabetic rats in a dose-dependent manner⁴⁰. Furthermore, *A. paniculata* extract was reported to reduce hyperglycemia by inhibiting β -cell dysfunction in alloxan-induced diabetic rats⁴¹. An ethanolic extract of *A. paniculata* and andrographolide lowered the plasma glucose levels by enhancing the translocation of glucose-transporter-4 in insulin-resistant obese mice ${}^{42, 43}$.

Effects on Cardiovascular Disease

An increase of blood- clotting time has been demonstrated by *A. paniculata*; thus making it useful for the pre- and post- treatments by extract of *A. paniculata* after surgery which prevent the contraction of blood vessels, hence decreasing the risk of the consequent closing of blood vessels after angioplasty operations⁴⁴. Several studies have used animal models to study the effects of aqueous extracts and active constituents of *A*.

paniculata, both before and after experimental myocardial infarction. An extract of the plant produced antihypertensive effects because it relaxed smooth muscles in the walls of blood vessels and prevented the blood vessels from constricting and limiting blood flow to the brain, heart, and other organs⁴⁵. It was demonstrated that the dichloromethane extract of *A. paniculata* significantly reduced coronary perfusion pressure by up to $24.5 \pm 3.0 \text{ mm Hg}$ at a 3 mg dose and also reduced the heart rate by up to $49.5 \pm 11.4 \text{ beats/min}$ at this dose⁴⁶. *A. paniculata* was also found to reduce the arterial constriction caused by high cholesterol in the diet and by injury to the inner lining of the blood vessel⁴⁷. The pretreatment of andrographolide; led to a time-dependent protection of rat cardiomyocytes against hypoxia injury which was reported to be caused by this effect was reported to be linked with regulation of cellular reduced glutathione (GSH) level and antioxidant enzyme activities⁴⁸. Dogs administered with *A. paniculata* one hour after the development of myocardial infarction were reported to have decreased damage of the heart muscle⁴⁹. These findings infer the promising use of *A. paniculata* as a favorable agent for cardiovascular therapy.

Hepatoprotective activity:

Among the phytochemicals obtained from the plant, andrographolide the main constituent of A. paniculata was found to be effective in preventing carbon tetrachloride induced liver damage (Rats and mice). Andrographolide has also exhibited significant hepatoprotective effect against different types of liver damages induced by paracetamole or galactsamine, ⁵⁰ and had a higher capacity than a classical antioxidant silymarin in preventing a decrease of bile production induced by paracetamole. ⁵¹ The paracetamol induced enhanced levels of serum marker enzymes such as serum glutamate pyruvate transaminase (GPT), serum glutamate oxaloacetate transaminase (GOT), alkaline phosphatase (ALP), and bilirubin in peripheral blood serum and distorted hepatic tissue architecture along with increased levels of lipid peroxides (LPO) and reduction of superoxide dismutase (SOD), catalase, reduced glutathione (GSH) and glutathione peroxidase (GPx) in liver tissue. Administration of the plant extracts after paracetamol insult restored the levels of these parameters to control (untreated) levels.⁵²

It also played a great hepatoprotective role by reducing a lipid peroxidation product malondialdehyde (MDA), in which the lowering of MDA formation conveyed the free radical scavenging property of diterpene lactones of andrographolide, as well as by maintaining high level of glutathione, glutamic pyruvate transminase, and alkaline phosphatase in carbontetrachloride or tert-butylhydroperoxide treated mice.⁵³ Antihepatotoxic action of andrographolide was reported against *Plasmodium berghei* K173-induced hepatic damage of mastomys natalensis.⁵⁴ Andrographolide was found to play a important role as effective stimulator of gall bladder function by producing a significant increase in bile flow, bile salt and bile acid in conscious rats and anesthetized guinea pigs. There was distinct improvement in the majority of infective hepatitis patient after continuous treatment with *A. paniculata* in regular recovery from jaundice, improvement of appetite and liver function tests, as well as subsidence of fever.⁵⁵

Conclusion: *A. paniculata* has been an important herb for treatment of various ailments in the traditional medicinal system. Plant extract of the plant contains various phytochemicals exhibiting promising pharmacological effects a few of them are discussed here.

Reference

1. Latto SK, Khan S, Dhar AK, Chaudhry DK, Gupta KK, Sharma PR. Genetics and mechanism of induced male sterility in Andrographis paniculata (Berm.f.) Nees and its significance. Curr Sci 2006; 91: 515-519.

- 2. L i J , Huang W , Z hang H , Wang X , Zhou H . Synthesis of andrographolide derivatives and their TNF -alpha and IL 6 expression inhibitory activities. Bioorg Med Chem Lett 2007; 17:6891-6894.
- 3. Mishra SK, Sangwan NS, Sangwan RS. Andrographis paniculata (Kalmegh): a review. Pharmacognosy Rev 2007; 1: 283-298.
- 4. Khare CP. Andrographis paniculata. In: Khare Khare CP, editor. Indian medicinal plants, an Illustrated Dictionary. New Delhi, India: Springer; 2007, p. 2, 49-50.
- 5. Chopra RN. Glossary of Indian medicinal plants. New Delhi: Council for Scientific and Industrial Research; 1980, p. 18.
- 6. Jarukamjorn K , Kondo S , Chatuphonprasert W , Sakuma T , Kawasaki Y , Emito N . Gender-associated modulation of inducible CYP1A1 expression by andrographolide in mouse liver. Eur J Pharm Sci 2010; 39: 394-401.
- 7. Chaturvedi GN, Tomar GS, T iwari SK, Singh KP. Clinical studies on Kalmegh (Andrographis paniculata Nees) in infective hepatitis. J Int Inst Ayurveda 1983; 2: 208-211.
- 8. Balu S, Alagesaboopathi C. Anti-inflammatory activities of some species of Andrographis Wall. Anc Sci Life 1993; 13: 180-184.
- 9. Saxena S , Jain DC , Bhakuni RS , Sharma RP . Chemistry and pharmacology of Andrographis species. Indian Drugs 1998 ; 35: 458-467.
- 10. Perry LM. Medicinal plants of East and Southeast Asia: attributed properties and uses. Cambridge: MIT Press; 1980.
- 11. Deng WL. Preliminary studies on the pharmacology of the Andrographis product dihydroandrographolide sodium succinate. Newslett Clin Herb Med 1978; 8: 26-28.
- 12. Alagesaboopathi C, Dwrakan P, Ramachandran VS. Andrographis paniculata Nee in tribal medicine of Tamil N adu. Anc Sci Life 1999; 19: 28-30.
- Panossian A, Davtyan T, Gukassyan N, Gukasova G, Mamikonyan G, G abrielian E, et al. E ffect of andrographolide and Kan Jang fixed combination of extract SHA - 10 and extract SHE - 3 on proliferation of human lymphocytes, production of cytokines and immune activation markers in blood cell culture. Phytomedicine 2002; 9: 598-605.
- 14. Poolsup N, Suthisisang C, Prathanturarug S, Asawamekin A, Chanchareon U. Andrographis paniculata in the symptomatic treatment of uncomplicated upper respiratory tract infection: systematic review of randomized controlled trials. J Clin Pharm Ther 2004; 29(1): 37-45.
- 15. Mishra N, Agrawal S, Jadhav SK, Kumar A; Traditional Applications and Phytochemical Investigation of *Andrographis paniculata* from Four Districts of Chhattisgarh, India Adv. Biores. 2014; 5 (2):172-182.
- 16. Phosphane N, Rangkadilok N, Thongnest S, Ruchirawat M, Ruchirawat J. Determination and variation of three active diterpenoids in Andrographis paniculata (Burm.f.) Nees. Phytochem Anal 2004; 15: 365-371.
- 17. L i WK , Fitzloff JF . HPLC PDA determination of bioactive diterpenoids from plant materials and commercial products of Andrographis paniculata. J Liq Chromatogr Relat Technol 2004; 27: 2407-2420.
- Siripong P, Kongkathip B, Preechanukool K, Picha P, Tunsuwan K, Taylor WC. Cytotoxic diterpenoid constituents from Andrographis paniculata Nees leaves. Sci Asia 1992; 18: 187-194.

- 19. Koteswara Rao Y, Vimalamma G, Rao CV, Tzeng YM. Flavonoids and andrographolides from Andrographis paniculata. Phytochemistry 2004; 65: 2317-2321.
- 20. Kuroyanagi M , Sato M , Ueno A , Nishi K . F lavonoids from Andrographis paniculata. Chem Pharm Bull 1987; 35: 4429-4435.
- Dua VK, Ojha VP, Roy R, Joshi BC, Valecha N, Devi CU, et al. Anti-malarial activity of some xanthones isolated from the roots of Andrographis paniculata. J Ethnopharmacol 2004; 95: 247-251.
- 22. Xu C, Chou GX, Wang CH, Wang ZT. Rare noriridoids from the roots of Andrographis paniculata. Phytochemistry 2012; 77: 275-279.
- 23. Singh PK, Roy S, Dey S. Antimicrobial activity of Andrographis paniculata. Fitoterapia 2003; 74: 692-694.
- Chandrasekaran CV, Gupta A, Agarwal A. Effect of an extract of Andrographis paniculata leaves on inflammatory and allergic mediators in vitro. J Ethnopharmacol 2010; 129: 203-207.
- 25. Parichatikanond W, Suthisisang C, Dhepakson P, Herunsalee A. Study of antiinflammatory activities of the pure compounds from Andrographis paniculata (Burm.f.) Nees and their effects on gene expression. Int Immunopharmacol 2010; 10: 1361-1373.
- 26. Abu-Ghefreh AA, Canatan H, Ezeamuzie C I. *In vitro* and *in vivo* anti-inflammatory effects of andrographolide. International Immunopharmacology Volume 9, Issue 3, March 2009, Pages 313-318.
- 27. Chandrasekaran CV, Thiyagarajan P, Deepak HB, Agarwal A. In vitro modulation of LPS /calcimycin induced inflammatory and allergic mediators by pure compounds of Andrographis paniculata (King of bitters) extract. Int Immunupharmacol 2011; 11: 70-84.
- 28. <u>P.P. Gupta</u>, <u>J.S. Tandon</u>, <u>G.K. Patnaik</u>, Antiallergic Activity of Andrographolides Isolated from Andrographis Paniculata (Burm. F) Wall <u>Pharmaceutical Biology</u> Volume 36, 1998 -<u>Issue 1</u>
- 29. Kumar RA, S ridevi K, K umar NV, N anduri S, R ajagopal S. Anticancer and immunostimulatory compounds from Andrographis paniculata. J Ethnopharmacol 2004; 92: 291-295.
- 30. Matsuda T, Kuroyanagi M, Sugiyama S, Umehara K, Ueno A, Nishi K . Cell differentiation inducing diterpenes from Andrographis paniculata Nees. Chem Pharm Bull 1994; 42: 1216-1225.
- 31. Nugroho AE, Andrie M, Warditiani NK, Siswanto E, Pramono S, Lukitaningsih E. Antidiabetic and antihiperlipidemic effect of Andrographis paniculata (Burm. f.) Nees and andrographolide in high-fructose-fat-fed rats. Indian J Pharmacol. 2012 May;44(3):377-81. doi: 10.4103/0253-7613.96343. PMID: 22701250; PMCID: PMC3371463.
- Dai Y, Chen SR, Chai L, Zhao J, Wang Y, Wang Y. Overview of pharmacological activities of *Andrographis paniculata* and its major compound andrographolide. Crit Rev Food Sci Nutr. 2019;59(sup1):S17-S29. doi: 10.1080/10408398.2018.1501657. Epub 2018 Sep 10. PMID: 30040451.
- 33. Adiguna SP, Panggabean JA, Atikana A, Untari F, Izzati F, Bayu A, Rosyidah A, Rahmawati SI, Putra MY. Antiviral Activities of Andrographolide and Its Derivatives: Mechanism of Action and Delivery System. Pharmaceuticals (Basel). 2021 Oct 28;14(11):1102. doi: 10.3390/ph14111102. PMID: 34832884; PMCID: PMC8619093.

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- 34. Hossain S, Urbi Z, Karuniawati H, Mohiuddin RB, Moh Qrimida A, Allzrag AMM, Ming LC, Pagano E, Capasso R. *Andrographis paniculata* (Burm. f.) Wall. ex Nees: An Updated Review of Phytochemistry, Antimicrobial Pharmacology, and Clinical Safety and Efficacy. Life (Basel). 2021 Apr 16;11(4):348. doi: 10.3390/life11040348. PMID: 33923529; PMCID: PMC8072717.
- 35. Ji X, Li C, Ou Y, Li N, Yuan K, Yang G, Chen X, Yang Z, Liu B, Cheung WW, Wang L, Huang R, Lan T. Andrographolide ameliorates diabetic nephropathy by attenuating hyperglycemia-mediated renal oxidative stress and inflammation via Akt/NF-κB pathway. Mol Cell Endocrinol. 2016 Dec 5;437:268-279. doi: 10.1016/j.mce.2016.06.029. Epub 2016 Jul 1. PMID: 27378149.
- 36. Islam MT. Andrographolide, a New Hope in the Prevention and Treatment of Metabolic Syndrome. Front Pharmacol. 2017 Aug 23;8:571. doi: 10.3389/fphar.2017.00571. PMID: 28878680; PMCID: PMC5572404.
- 37. Naik RR, Munipally PK, Nagaraju T. Andrographolide reorganise hyperglycaemia and distorted antioxidant profile in streptozotocin-induced diabetic rats. Cardiovasc Hematol Agents Med Chem. 2017 Oct 26. doi: 10.2174/1871525715666171026115248. Epub ahead of print. PMID: 29076435.
- Mehta S., Sharma A. K., Singh R. K. (2021). Pharmacological activities and molecular mechanisms of pure and crude extract of Andrographis paniculata: An update. *Phytomedicine Plus* 1 (4), 100085. 10.1016/j.phyplu.2021.100085
- Syukri Y, Taher M, Martien R, Lukitaningsih E, Nugroho AE, Zakaria ZA. Selfnanoemulsifying Delivery of Andrographolide: Ameliorating Islet Beta Cells and Inhibiting Adipocyte Differentiation. Adv Pharm Bull. 2021 Jan;11(1):171-180. doi: 10.34172/apb.2021.018. Epub 2020 Nov 7. PMID: 33747864; PMCID: PMC7961231.
- 40. Xu FF, Fu SJ, Gu SP, Wang ZM, Wang ZZ, He X, Xiao W. Simultaneous determination of andrographolide, dehydroandrographolide and neoandrographolide in dog plasma by LC-MS/MS and its application to a dog pharmacokinetic study of Andrographis paniculata tablet. J Chromatogr B Analyt Technol Biomed Life Sci. 2015 May 15;990:125-31. doi: 10.1016/j.jchromb.2015.03.014. Epub 2015 Mar 30. PMID: 25864014.
- 41. Jaiyesimi KF, Agunbiade OS, Ajiboye BO, Afolabi OB. Polyphenolic-rich extracts of *Andrographis paniculata* mitigate hyperglycemia via attenuating β-cell dysfunction, proinflammatory cytokines and oxidative stress in alloxan-induced diabetic Wistar albino rat. J Diabetes Metab Disord. 2020 Nov 15;19(2):1543-1556. doi: 10.1007/s40200-020-00690-2. PMID: 33553038; PMCID: PMC7843849.
- 42. Akhtar MT, Bin Mohd Sarib MS, Ismail IS, Abas F, Ismail A, Lajis NH, Shaari K. Anti-Diabetic Activity and Metabolic Changes Induced by Andrographis paniculata Plant Extract in Obese Diabetic Rats. Molecules. 2016 Aug 9;21(8):1026. doi: 10.3390/molecules21081026. PMID: 27517894; PMCID: PMC6273188.
- 43. Chen CC, Lii CK, Lin YH, Shie PH, Yang YC, Huang CS, Chen HW. Andrographis paniculata Improves Insulin Resistance in High-Fat Diet-Induced Obese Mice and TNFα-Treated 3T3-L1 Adipocytes. Am J Chin Med. 2020;48(5):1073-1090. doi: 10.1142/S0192415X20500524. PMID: 32668968.

- 44. Wang HW, Zhao HY, Xiang SQ. Effects of *Andrographis paniculata* component on nitric oxide, endothelin and lipid peroxidation in experimental atherosclerotic rabbits. *Zhongguo Zhong Xi Yi Jie He Za Zhi*. 1997;17(9):547–549.
- 45. Huang LY. The effects of andrographolide on experimental blood deficiency of cardiac muscle. *Chinese Herbal Medicine*. 1987;18:26–28.
- 46. Awang K, Abdullah NH, Hadi AH, Fong YS. Cardiovascular activity of labdane diterpenes from *Andrographis paniculata* in isolated rat hearts. *Journal of Biomedicine and Biotechnology*. 2012;2012:5 pages.876458
- 47. Wang D, Zhao H. Experimental studies on prevention of atherosclerotic arterial stenosis and restenosis after angioplasty with *Andrographis paniculata* Nees and Fish Oil. *Journal of Tongji Medical University*. 1993;13(4):193–198.
- Woo AYH, Waye MMY, Tsui SKW, Yeung STW, Cheng CHK. Andrographolide upregulates cellular-reduced glutathione level and protects cardiomyocytes against hypoxia/reoxygenation injury. *The Journal of Pharmacology and Experimental Therapeutics*. 2008;325(1):226–235.
- Zhao H, Fang W. Protective effects of Andrographis paniculata Nees on post-infarction myocardium in experimental dogs. Journal of Tongji Medical University. 1990;10(4):212– 217.
- 50. 50. Handa SS, Sharma A. Hepatoprotective activity of andrographolide against galoctosomine and paracetamole intoxication in rats. Indian J Med Res. 1990; 92: 284-292.
- 51. Handa SS, Sharma A. Hepatoprotective activity of andrographolide from Andrographis paniculata against carbontetrachloride. Indian J Med Res. 1990; 92: 276-283.
- 52. Nagalekshmi R., Menon A., Chandrasekharan DK, Nair CKK, Hepatoprotective activity of Andrographis Paniculata and Swertia Chirayita, Food and Chemical Toxicology, Volume 49, Issue 12, 2011, Pages 3367-3373, ISSN 0278-6915, <u>https://doi.org/10.1016/j.fct.2011.09.026</u>
- 53. Kapil A, koul IB, Banerjee SK, Gupta BD. Antihepatotoxic effect of major diterpenoid constituents of Andrographis paniculata Biochem. Pharmacol. 1993; 46: 182-185.
- 54. Chander R, Srivastava V, Tandon JS, Kapoor NK. Antihepatotoxic activity of diterpenes of *Andrographis paniculata* (Kalmegh) against plasmodium berghei-induced hepatic damage in mastomys Natalensis. Pharm Biol, 1995; 33: 135-138.
- 55. Shukla B, Visen PKS, Dhawan BN. Choleretic effect of andrgrapholide in rats and guinea pigs. Planta Med. 1992; 58: 146-148.

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